



Published in final edited form as:

Cancer Discov. 2012 December ; 2(12): 1091-1099. doi:10.1158/2159-8290.CD-12-0329.

The initial hours of metastasis: the importance of cooperative host-tumor cell interactions during hematogenous dissemination

Myriam Labelle and Richard O. Hynes

Howard Hughes Medical Institute, Koch Institute for Integrative Cancer Research, Massachusetts Institute of Technology, Cambridge, MA 02139, USA

Abstract

Tumor cells transit from the primary tumor via the blood circulation to form metastases in distant organs. During this process, tumor cells encounter a number of environmental challenges and stimuli that profoundly impact their metastatic potential. Here, we review the cooperative and dynamic host-tumor cell interactions that support and promote the hematogenous dissemination of cancer cells to sites of distant metastasis. In particular, we discuss what is known about the crosstalk occurring among tumor cells, platelets, leukocytes and endothelial cells and how these cell-cell interactions are organized both temporally and spatially at sites of extravasation and in the early metastatic niche.

Keywords

Metastatic niche; host-tumor cell interactions; platelets; leukocytes; extravasation

Introduction

Metastasis is the cause of about 90% of cancer-associated deaths, yet the mechanisms governing this clinically important process remain poorly understood. Tumor cells can metastasize via the lymphatics to neighboring lymph nodes. However, it remains unclear, in the general case, whether lymph nodes serve as a "way-station" en route to the vasculature. Distant metastases rely on hematogenous dissemination via the blood circulation and we will concentrate here on this latter process. In order to metastasize successfully, cancer cells must complete several complex sequential steps: detachment from the primary tumor, intravasation into the vascular system (whether directly or via lymphatics and lymph nodes), survival while in transit through the circulation, initial arrest, extravasation, initial seeding, and survival and proliferation in the target tissue. Despite the fact that large primary tumors can shed millions of cells into the vasculature every day, very few metastases eventually develop (1, 2). Thus, metastasis is, overall, an inefficient process, implying that tumor cells frequently fail to execute one or more of the required steps of the metastatic cascade. Tumor cells that succeed in forming metastases may have acquired the necessary traits to complete these steps while still in the primary tumor, either autonomously or as a result of changes induced by inflammation, stromal cells or other environmental conditions (e.g., hypoxia, mechanical forces) present in the primary tumor (3). However, the metastatic potential of tumor cells is also further very significantly modulated by the environmental conditions and

Correspondence: Richard Hynes, Massachusetts Institute of Technology, 77 Massachusetts Ave, 76-361D, Cambridge, MA 02139, Tel: 617-253-6422, Fax: 617-253-8357, rohynes@mit.edu.

The authors disclose no potential conflicts of interest.

NIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH-PA Author Manuscript

READ

BIG CONCEPT

host cells, in particular platelets and bone marrow-derived cells (BMDCs) that tumor cells encounter during their transit through the bloodstream and at the sites of distant metastases. This aspect of the metastatic cascade remains poorly understood, due to the technical challenges associated with imaging, isolation and analysis of circulating tumor cells (CTCs) or single disseminated tumor cells (DTCs) that have metastasized to distant organs.

Nevertheless, recent studies using experimental mouse models have begun to demonstrate the importance of host-tumor cell interactions, both in the circulation and at sites of extravasation, for the establishment of metastasis. Many of these studies have been conducted with intravenous injections of tumor cells (experimental metastasis), which is generally considered a standard model for studying hematogenous dissemination. While this experimental setup presents some limitations (e.g. absence of a primary tumor, injection of tumor cells in a single event rather than scattered over a long period of time), it also offers important experimental advantages: it allows close temporal monitoring of the early interactions between single tumor cells and the host microenvironment and a precise characterization of the specific steps of the metastatic cascade affected by a given experimental treatment (4).

In this review, we discuss the sequence of events and key host cell types that interact with tumor cells during their hematogenous transit and their initial establishment at the secondary site and how these interactions influence metastasis and cancer prognosis.

Transit Through the Bloodstream and Initial Arrest (First Minutes)

Circulating tumor cells (CTCs) are frequently found in the blood of patients with primary solid tumors, and it is generally assumed that a subset of these cells will eventually give rise to distant metastases (5, 6). However, as indicated by intravascular injection of tumor cells into animal models, CTCs typically do not spend much time circulating through the bloodstream. Indeed, most carcinoma cells have diameters that are too large to pass through small capillaries and many are therefore trapped in the first capillary bed that they encounter within minutes of entering the circulation (Figure 1, 2A) (2). During this short period of transit, as well as during initial arrest, cells remain exposed to the blood flow and are vulnerable to death induced by shear stress and turbulence or by immune cells, particularly natural killer (NK) cells. Thus, tumor cells that have intrinsic traits enabling them to escape immune surveillance or to interact with shielding host cells would have an increased rate of success in this early phase of the metastatic cascade.

In this respect, activation of the coagulation cascade and the formation of platelet-rich thrombi around tumor cells in the vasculature have both been proposed to play major roles in physically shielding CTCs from the stress of blood flow and from lysis by natural killer (NK) cells (Figure 2A) (7-11). Tissue factor (TF) expressed by tumor cells triggers the formation of thrombin, which leads to both coagulation and platelet activation. In turn, coagulation and platelet activation enhance metastatic spread (11-16). Fibrin can be bound by integrins, $\alpha v \beta 3$ on tumor cells and $\alpha IIb \beta 3$ on activated platelets, leading to formation of tumor cell-fibrin-platelet aggregates. Therefore, genetic elimination or blockade of $\beta 3$ integrins on platelets or tumor cells compromises metastasis (8-10, 17-19). Furthermore, expression of diverse P-selectin ligands by tumor cells also favors their interaction with P-selectin on the surfaces of activated platelets (20). Consequently, platelet-tumor cell microthrombi do not form in P-selectin^{-/-} mice and these mice are protected from metastasis (21, 22). More generally, diverse intact platelet functions (e.g. platelet activation, adhesion via surface receptors, TGF β release) have been shown to be necessary for efficient metastasis in many experimental models (10, 17, 18, 23-26). Platelets are in fact, the major source of TGF β in the circulation (23). Consistent with a pro-metastatic role of platelets,

NIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH-PA Author Manuscript

1 2 2
BIG CONCEPTS

which needs Fibrin

Fibrin

high platelet counts and coagulation correlate with decreased patient survival in many different types of cancers, although some of that correlation could well be due to thrombosis caused by the presence of a tumor (9, 10, 27, 28).

In addition to providing physical shielding, platelets and coagulation have been shown to impair NK cell tumorolytic activity in vitro (7, 8). For example, platelet-derived TGF β downregulates the activating immunoreceptor NKG2D on NK cells (29) and PDGF released by platelets can also suppress NK cell function (30). The coating of the surfaces of tumor cells with normal platelet-derived MHC class I may also favor tumor cell escape from the innate immune system (31). Thus multiple platelet-tumor cell interactions can lead to the inhibition of NK cells, leading to increased tumor cell survival in the circulation.

Clustering of tumor cells and adhesion with other cell types have also been proposed to contribute to successful tumor cell survival in the circulation. For example, CTC clusters isolated from the blood of patients with metastatic prostate cancer have higher hematoxylin and eosin staining intensity than do individual CTCs, suggesting reduced cell death and potential protection from shear stress (6, 32). Similarly, CTCs incorporated in heterotypic tumor-fibroblast aggregates retrieved from the blood of tumor-bearing mice have improved viability compared with single CTCs (33). Given the enhancing effects of platelets on metastasis, it is plausible that the CTCs that are most effective in metastasis will prove to be those in aggregates with platelets and possibly also leukocytes. Most current methods for scoring CTCs in patients score only single cells and could be missing an important fraction of the CTC population.

Arrest and Adhesion to the Vascular Wall (First Hours)

The propensity of tumor cells to metastasize to specific organs is in part dependent on the circulation pattern, and the preferred sites of metastasis for a given type of cancer often include the first capillary beds downstream of the primary tumor. Examples are metastasis of colon cancer cells to the liver and of breast cancer cells to the lungs, where the initial arrest of tumor cells may be mainly caused by physical restriction in capillaries of small diameter (2). In such cases, the formation of aggregates comprising CTCs and host cells may enhance passive trapping in capillaries by increasing the diameter of tumor cell emboli. However, during metastasis to either the liver or the lung, tumor cells can also arrest in vessels of larger diameter than capillaries (34), demonstrating that active adhesion to the vasculature via specific proteins, such as selectins, integrins and metadherin, can also contribute to initial arrest (19, 35–38). Importantly, some of these adhesion receptors could be contributed by associated platelets, leukocytes or stromal fibroblasts.

It is likely that initial trapping, which occurs within minutes of the entry of tumor cells into the circulation, is mostly passive and dependent on circulation patterns, while the cells that permanently arrest are those that form specific, longer-lasting adhesive interactions with the endothelium. In accordance with this concept, a high proportion of tumor cells rapidly arrests in capillaries in experimental metastasis models, while sustained adhesion to the endothelium leading to permanent seeding seems to be of variable efficiency and often fails. Indeed, while some studies have shown that more than 80% of tumor cells survive the circulatory phase of metastasis and successfully seed the lungs after 24 hours (39), others have reported much lower rates of cell retention (~20% or less) at this same time point (1, 40–42). Thus, the rate of tumor cell death or displacement to other organs at this early stage of metastasis can, in many cases, be very high. For example, using real-time imaging *in vivo*, Kienast et al. observed that melanoma or lung carcinoma cells initially arrested in brain capillaries can enter and leave their arrested positions several times during the first 24 hours after intravascular injection (43). A high proportion of these cells die or are dislodged,

immune #1
platelets

while some others stably adhere to the endothelium and extravasate. The choices between displacement or retention at the initial site of arrest and subsequent extravasation may depend on specific traits of tumor cells or on influences of their associated thrombi, platelets and leukocytes. Only tumor cells that have extravasated but are residing in close apposition to blood vessels are eventually able to form overt metastases, suggesting that this specific microenvironment provides cells with prosurvival factors. Furthermore, tumor cells were found to home preferentially to discrete foci of vascular hyperpermeability in lungs (44). Soluble factors secreted by primary tumors increase the formation of hyperpermeable foci via localized activation of endothelial FAK and E-selectin, which in turn favors the adhesion of tumor cells to the endothelium (44). In addition, soluble factors secreted by primary tumors have been reported to induce the recruitment of BMDCs to specific areas of distant organs to form so-called premetastatic niches. These niches have been proposed to create a supportive environment for the survival and growth of incoming tumor cells (45–49). The presence of a primary tumor also triggers inflammation, which leads to the activation of the endothelium and platelets and contributes to the systemic mobilization of various types of BMDCs (immature myeloid cells, neutrophils, monocytes), which may all play critical and concerted roles in metastasis (3, 42, 49–51).

The presence of an activated endothelium may favor arrest and adhesion of tumor cells and this likely involves participation of myeloid cells (Figure 2B). For example, activation of the endothelium by IL-1 α , IL-1 β or TNF- α leads to the expression of E-selectin and P-selectin as well as VCAM-1 and ICAM-1 at the surfaces of endothelial cells. Binding of these cell adhesion molecules to their ligands on tumor cells can then promote tumor cell rolling and adhesion (20, 52, 53). Interestingly, in a liver metastasis model, the presence of tumor cells triggers the production of TNF- α by Kupffer cells, demonstrating that immune cells can play an active part in endothelial cell activation and therefore in favoring metastatic arrest (54). Similarly, Laubli et al. showed that activation of the endothelium *in vitro* by tumor cells is P-selectin-dependent and requires the simultaneous presence of platelets and neutrophils together with tumor cells (55). In addition to favoring tumor cell adhesion, the activated endothelium secretes the inflammatory cytokine CCL5, which promotes the recruitment of monocytes in proximity to the tumor cells. Importantly, platelets also secrete high levels of a plethora of growth factors and cytokines (e.g. PDGF, TGF β , PF4/CXCL4, VEGF, SDF-1, CXCL7, CCL5), which could also contribute to endothelial activation or directly lead to the recruitment of BMDCs (56). Furthermore, the presence of P-selectin on activated platelets adherent to the endothelium enhances the recruitment of leukocytes via binding to P-selectin glycoprotein ligand-1 (PSGL-1) on leukocytes (57–59). This interaction promotes the activation of the leukocyte β 2 integrin, which then binds to fibrinogen presented by α IIb β 3 integrin and to GPIIb α , ICAM-2 and JAM-3, all present on the surfaces of platelets and thereby stabilizes platelet-leukocyte interactions (60–63). Thus, the formation of cellular assemblies composed of tumor cells, platelets, leukocytes and activated endothelium appear very likely to be required for efficient metastasis.

Although the contributions of leukocytes to the primary tumor are well established, their roles in the processes of metastasis have been less well characterized until recently. Globally, leukocytes have been shown to support the early stages of metastasis, as illustrated by the decrease in leukocyte-tumor cell interactions and impaired early tumor cell seeding in L-selectin $^{-/-}$ mice (64). Similarly, metastasis was attenuated in mice unable to induce L-selectin ligand expression at sites of intravascular tumor cell arrest (40). Moreover, metastasis is reduced by genetic or pharmacological ablation of monocyte/macrophage-lineage cells (42, 51, 65), and tail-vein injection of neutrophils 1 hour after injection of melanoma cells results in increased retention of tumor cells in the lungs after 24 hours (66) (Figure 1, 2B). These latter observations may be explained by the secretion of IL-8 by tumor cells, which can attract and activate neutrophils by increasing their expression of β 2

integrins and adhesion to tumor cells (66). In turn, MMP-9 produced by neutrophils promotes the early survival of metastatic cells (6–24 hours) but has no effect on subsequent metastatic growth (67). On the other hand, Granot et al. recently showed that tumor-entrained neutrophils (TENs; a subset of CD11b⁺Ly-6G⁺MMP-9⁺ neutrophils isolated from tumor-bearing mice) can counteract metastatic seeding of breast carcinoma cells in the lungs by killing tumor cells via the generation of high levels of hydrogen peroxide (68). These anti-metastatic effects were observed upon the transfer of TENs into mice, but not if G-CSF-stimulated neutrophils were used. Thus, neutrophils can either promote or inhibit metastasis, depending on the stimuli to which they are exposed. Presumably, the presence of other host cells and factors determines the outcome of neutrophil-tumor interactions. For example, the killing activity of TENs can be blocked by TGF β in vitro, suggesting that a TGF β -rich microenvironment (such as that produced by platelet aggregation with tumor cells (23) could impede the function of TENs in vivo and promote metastasis, similarly to the context-dependent activity of neutrophils observed in primary tumors (69).

Extravasation and initial seeding (First Days)

Extravasation efficiency and kinetics depend both on tumor cells' intrinsic behavior and host tissue characteristics, and tumor cells that can extravasate rapidly presumably have an advantage during the metastatic cascade due to their ability to escape promptly from the hostile environment of the blood flow (70, 71). Indeed, cancer cells that are prone to metastasize to the lungs express high levels of Angptl4 or VEGF-A, two secreted factors that disrupt endothelial cell-cell junctions and facilitate extravasation (72, 73). Similarly, upregulation of other genes involved in vascular and extracellular matrix remodeling (EREG, COX2, MMP1 and MMP2) promotes extravasation and metastasis (70).

Extravasation, which typically occurs within 1 to 3 days (Figure 1), can also be directly enhanced by platelet-tumor cell interactions, once tumor cells enter the bloodstream. Mechanistically, platelet-derived TGF β and direct platelet-tumor cell contacts synergistically activate the TGF β /Smad and NF- κ B pathways in cancer cells, inducing an epithelial-mesenchymal transition in the tumor cells in vitro, and enhancing their extravasation and seeding in vivo (Figure 2C) (23). Platelet-specific ablation of TGF β 1 leads to reduced metastasis and to the impairment of tumor cell extravasation, directly demonstrating the requirement for platelet-derived TGF β in this process. Platelet-activated tumor cells also acquire a prometastatic gene expression signature, which includes enhanced expression of various proteases, cytokines and growth factors (23) that may contribute to metastasis not only by directly enhancing tumor cell invasive potential but also by modifying the microenvironment. Importantly, these results reveal that platelets are more than physical shields and that the metastatic potential of tumor cells continues to evolve outside the primary tumor site, in response to their interactions with platelets in the bloodstream. Therefore, by triggering the activation of specific signaling pathways in tumor cells, platelets may initiate a cascade of events reaching beyond the initial hours of metastasis and impacting subsequent steps of the metastasis cascade, such as survival and growth at the secondary site. For example, activation of the NF- κ B pathway in tumor cells in response to interaction with platelets promotes the expression of CCL2, a proinflammatory chemokine involved in monocyte recruitment (23, 74). In experimental metastasis models, CCL2 secretion by both tumor cells and stromal cells was shown to recruit inflammatory monocytes to the lungs early after the injection of breast tumor cells (Figure 2C) (51, 75). Tissue factor (TF) produced by the tumor cells also enhances coagulation and this too contributes to attract myeloid cells to the vicinity of tumor cells (65). The monocyte/macrophage-lineage cells recruited by the tumor cells were shown to enhance the seeding of metastatic mammary tumor cells in the lung (42, 51, 65). Among these cell populations, a distinct set of metastasis-associated macrophages (F4/80⁺, CD11b⁺,

Gr1⁺) secrete VEGF-A that promotes the extravasation, seeding and growth of the tumor cells (42), presumably via increased endothelial permeability. However, in a colon carcinoma experimental metastasis model, tumor cell-derived CCL2 has also been shown to signal directly to CCR2 expressed by endothelial cells, resulting in an increase in vascular permeability and subsequent metastasis by a mechanism independent of myeloid cells (Figure 2C) (75). Thus, tumor cell extravasation is facilitated by multiple complex interactive networks comprising direct platelet-to-tumor cell signaling, tumor cell-to-endothelium signaling and monocyte/macrophage-to-endothelium signaling.

Another example of a prometastatic cascade of events involving multiple types of host cells was provided by Laubli et al. (55), who showed that colon carcinoma cells together with platelets and neutrophils activate the endothelium. In turn, the activated endothelial cells secrete CCL5, which leads to increased recruitment of monocytes to the tumor cells. In this model, monocyte recruitment occurs after 2 days (55), a time point at which platelets are no longer associated with tumor cells, illustrating the sequential involvement of different host cells in supporting metastatic seeding. Although not yet tested, it is a plausible hypothesis that early and transient platelet-tumor cell interactions trigger a cascade of paracrine signals impinging on the recruitment and function of various types of leukocytes, which in turn contribute to successful survival and metastasis. Indeed, macrophages and specific subsets of bone-marrow-derived immature cells have been implicated in promoting cell survival and proliferation in models of metastasis to the lungs. For example, binding of VCAM-1 aberrantly expressed by tumor cells to $\alpha 4$ integrin expressed by macrophages, protects cancer cells from proapoptotic cytokines such as TRAIL, leading to increased survival and metastasis (76). Other examples of the importance of tumor cell-stroma interactions for early metastatic colonization come from recent studies, which demonstrated requirements for periostin and tenascin C expression by fibroblasts at the site of metastasis for successful metastatic growth (Figure 2C) (77, 78). TGF β seems to be involved in the enhancement of the expression of these two ECM proteins, suggesting that TGF β expressed by tumor cells or host cells (such as platelets, as discussed above) may be important for the initiation of this supportive metastatic niche (78–80). Finally, it is likely that the tumor-promoting effects of BMDCs, which are increasingly well understood for tumor progression at the primary site, may also be important for the subsequent establishment of overt metastases.

The first hours of metastasis as a possible therapeutic target

Most of the approved anti-cancer therapies inhibit the growth of primary tumors. While some of those therapies also have an effect on metastatic growth, there are currently no therapies specifically aimed at preventing the metastatic process by targeting the different steps of the metastatic cascade. Furthermore, while some potential anti-metastatic compounds have been identified in preclinical models, there is a clear need for clinical trials specifically designed to test for anti-metastatic effects (e.g. time required for the formation of a new metastasis) rather than for the ability of compounds to prevent tumor growth (81).

The early steps of the metastatic cascade discussed in this review are generally not considered as attractive clinical targets. The rationale for this opinion is that tumor cells can disseminate early during tumor progression (82, 83), and therefore it is likely that some metastatic cancer cells have already completed the early steps of the metastatic cascade by the time of cancer diagnosis. Thus, the later steps comprising escape from dormancy, reinitiation of growth, colonization and survival in the metastatic niche are likely better targets for therapeutic intervention. Indeed, while CTCs can complete the steps of the metastatic cascade leading to seeding within a few days, reinitiation of growth can be significantly delayed and metastatic growth occurs over an extended period of time, providing a more manageable time window for therapeutic intervention.

That said, the early steps of metastatic dissemination discussed here may offer some new opportunities for therapeutic interventions targeting molecular mechanisms and cellular processes such as adhesion, migration, invasion and epithelial-to-mesenchymal transition, that are not affected by the cytotoxic or antiproliferative effects of most traditional anti-cancer therapies. In addition, cells transiting through the bloodstream may be particularly accessible to pharmacological intervention. Drugs or combinations of drugs impairing not only the ability of tumor cells to proliferate, but also their ability to interact with host cells and complete the early steps of the metastatic cascade may prove beneficial to prevent further metastatic dissemination either from the primary tumor or from already existing metastases. Indeed, it has been demonstrated in animal models that tumor cells from metastases have the ability to re-enter the circulation and seed other metastases (84, 85) or to self-seed back at the primary tumor site, further contributing to cancer progression (86). Thus, inhibitors of metastatic arrest, extravasation or seeding may impact overall disease progression, even if disseminated tumor cells are already present in a patient. However, the patients most likely to benefit from metastatic prevention therapy would be those who have been diagnosed at early stages prior to detectable metastatic spread, or even people that do not have the disease but are at high risk for developing highly metastatic cancers. Specific inhibitors of metastasis could also be envisaged for cases where surgical ablation of the primary tumor is not possible, or during the perioperative period, since surgery may enhance the release of CTCs into the bloodstream (87, 88).

Even though many aspects of the early steps of metastasis are still incompletely understood, molecules that are critical for the completion of specific steps of the metastatic cascade are starting to emerge and could possibly be exploited as therapeutic targets. Inhibitors of a few of the signaling pathways involved in the early steps of metastasis (e.g. VEGF-A, TGF β , NF- κ B, CCL2) are already used in the clinic or are being evaluated in clinical trials for the treatment of cancers or other diseases. Furthermore, interfering with the ability of tumor cells to recruit or interact with supportive host cells may prevent the formation of optimal conditions for metastatic progression. In this respect, inhibitors of cell adhesion receptors required for the tumor cell-host cell interactions may be of particular interest.

For example, the anticoagulants heparin or low molecular weight heparin (LMWH) have already been shown to prevent metastasis in preclinical models by inhibiting the formation of platelet-tumor cell aggregates (22). More importantly, a number of independent clinical trials have demonstrated that treatment with LMWH improves the survival of cancer patients (9, 89–91). Interestingly, the beneficial effects of a LMWH treatment were predominantly seen in patients with good prognosis or that did not have detectable metastasis at the onset of treatment, consistent with a role for LMWH in inhibiting the seeding of metastases rather than in the growth of existing ones (9, 89–91). Mechanistically, the effect of heparin on metastasis is attributed primarily to its ability to inhibit the interaction of P-selectin with its ligands and not to its anticoagulant activity. Indeed, the synthetic LMWH fondaparinux, which does not inhibit P-selectin but retains anticoagulant activity, fails to inhibit experimental metastasis (92).

In addition to inhibiting P-selectin, heparin can also inhibit L-selectin, and α 4 β 1 and α IIb β 3 integrins (93), providing indications that heparin might interfere with multiple pro-metastatic adhesive interactions. Similarly, function-blocking antibodies targeting α 4 β 1 or α IIb β 3 integrins inhibit experimental metastasis (94, 95). Moreover, small molecules and antibodies that inhibit the function of α IIb β 3 integrin or the binding of VCAM-1 to α 4 integrin are already available in the clinic. Antagonists of α IIb β 3 integrin are used as anti-thrombotics and could be expected to block platelet-tumor cell interactions through fibrinogen as would also be true for antagonists of α v β 3 integrin, present on many tumor cells. Antagonists of α 4 and β 2 integrins have been developed for the treatment of diseases

involving influx of leukocytes, such as inflammation and autoimmunity, and might thus also interfere with the macrophage-tumor cell interactions that promote tumor cell arrest, survival and reinitiation of tumor growth at the site of metastasis (96, 97).

Overall, while inhibitors of host-tumor cell signaling interactions show promise in experimental models, it remains to be tested whether these agents will prevent or significantly delay metastasis in cancer patients, and clinical trials will be challenging given the long time scales necessary. Furthermore, potential side effects affecting vital functions such as the immune response, coagulation and hemostasis will need to be carefully evaluated. Thus, a better and more comprehensive understanding of the molecular mechanisms involved in metastasis is required for the development of specific therapies with minimal potential adverse effects and efficient blocking of cancer metastasis.

Conclusions

While the complexity of the metastatic cascade has been acknowledged for many years, the active participation of cells of the host microenvironment to metastatic dissemination is only beginning to be appreciated. The studies reviewed above provide examples of the importance of dynamic tumor-host cell interactions at each step of the metastatic cascade (Figures 1 and 2). The context-dependent and concerted actions of different populations of host cells appear to be necessary for efficient metastasis. However, exactly how the different types of host cells interact with each other as well as with tumor cells both temporally and spatially, and the precise hierarchy and function of these interactions remain incompletely understood. Answering these fundamental questions will likely provide important clues not only about the molecular mechanisms involved during metastatic dissemination but also about how these early processes influence the subsequent metastatic colonization. Deeper understanding of these diverse tumor-host cell interactions may also offer possibilities for novel therapeutic interventions.

Acknowledgments

Financial support: The work in the authors' laboratory was supported by the National Cancer Institute (U54 CA126515), the Ludwig Center at MIT, the Koch Institute and the Howard Hughes Medical Institute of which ROH is an Investigator. ML was supported by an Anna Fuller Postdoctoral Fellowship.

References

1. Fidler IJ. Metastasis: quantitative analysis of distribution and fate of tumor emboli labeled with ¹²⁵I-5-iodo-2'-deoxyuridine. *J Natl Cancer Inst.* 1970; 45:773-82. [PubMed: 5513503]
2. Chambers AF, Groom AC, MacDonald IC. Dissemination and growth of cancer cells in metastatic sites. *Nat Rev Cancer.* 2002; 2:563-72. [PubMed: 12154349]
3. Joyce JA, Pollard JW. Microenvironmental regulation of metastasis. *Nat Rev Cancer.* 2009; 9:239-52. [PubMed: 19279573]
4. Khanna C, Hunter K. Modeling metastasis in vivo. *Carcinogenesis.* 2005; 26:513-23. [PubMed: 15358632]
5. Cristofanilli M, Budd GT, Ellis MJ, Stopeck A, Matera J, Miller MC, et al. Circulating tumor cells, disease progression, and survival in metastatic breast cancer. *N Engl J Med.* 2004; 351:781-91. [PubMed: 15317891]
6. Yu M, Stott S, Toner M, Maheswaran S, Haber DA. Circulating tumor cells: approaches to isolation and characterization. *J Cell Biol.* 2011; 192:373-82. [PubMed: 21300848]
7. Nieswandt B, Hafner M, Echtenacher B, Mannel DN. Lysis of tumor cells by natural killer cells in mice is impeded by platelets. *Cancer Res.* 1999; 59:1295-300. [PubMed: 10096562]

8. Palumbo JS, Talmage KE, Massari JV, La Jeunesse CM, Flick MJ, Kombrinck KW, et al. Platelets and fibrin(ogen) increase metastatic potential by impeding natural killer cell-mediated elimination of tumor cells. *Blood*. 2005; 105:178–85. [PubMed: 15367435]
9. Erpenbeck L, Schon MP. Deadly allies: the fatal interplay between platelets and metastasizing cancer cells. *Blood*. 2010; 115:3427–36. [PubMed: 20194899]
10. Gay LJ, Felding-Habermann B. Contribution of platelets to tumour metastasis. *Nat Rev Cancer*. 2011; 11:123–34. [PubMed: 21258396]
11. Degen JL, Palumbo JS. Hemostatic factors, innate immunity and malignancy. *Thromb Res*. 2012; 129 (Suppl 1):S1–5. [PubMed: 22682116]
12. Mueller BM, Reisfeld RA, Edgington TS, Ruf W. Expression of tissue factor by melanoma cells promotes efficient hematogenous metastasis. *Proc Natl Acad Sci U S A*. 1992; 89:11832–6. [PubMed: 1465406]
13. Palumbo JS, Kombrinck KW, Drew AF, Grimes TS, Kiser JH, Degen JL, et al. Fibrinogen is an important determinant of the metastatic potential of circulating tumor cells. *Blood*. 2000; 96:3302–9. [PubMed: 11071621]
14. Palumbo JS, Potter JM, Kaplan LS, Talmage K, Jackson DG, Degen JL. Spontaneous hematogenous and lymphatic metastasis, but not primary tumor growth or angiogenesis, is diminished in fibrinogen-deficient mice. *Cancer Res*. 2002; 62:6966–72. [PubMed: 12460914]
15. Im JH, Fu W, Wang H, Bhatia SK, Hammer DA, Kowalska MA, et al. Coagulation facilitates tumor cell spreading in the pulmonary vasculature during early metastatic colony formation. *Cancer Res*. 2004; 64:8613–9. [PubMed: 15574768]
16. Liu Y, Jiang P, Capkova K, Xue D, Ye L, Sinha SC, et al. Tissue factor-activated coagulation cascade in the tumor microenvironment is critical for tumor progression and an effective target for therapy. *Cancer Res*. 2011; 71:6492–502. [PubMed: 21880589]
17. Karpatkin S, Pearlstein E, Ambrogio C, Collier BS. Role of adhesive proteins in platelet tumor interaction in vitro and metastasis formation in vivo. *J Clin Invest*. 1988; 81:1012–9. [PubMed: 3280598]
18. Nierodzick ML, Plotkin A, Kajumo F, Karpatkin S. Thrombin stimulates tumor-platelet adhesion in vitro and metastasis in vivo. *J Clin Invest*. 1991; 87:229–36. [PubMed: 1845869]
19. Felding-Habermann B, O'Toole TE, Smith JW, Fransvea E, Ruggeri ZM, Ginsberg MH, et al. Integrin activation controls metastasis in human breast cancer. *Proc Natl Acad Sci U S A*. 2001; 98:1853–8. [PubMed: 11172040]
20. Kim YJ, Borsig L, Han HL, Varki NM, Varki A. Distinct selectin ligands on colon carcinoma mucins can mediate pathological interactions among platelets, leukocytes, and endothelium. *Am J Pathol*. 1999; 155:461–72. [PubMed: 10433939]
21. Kim YJ, Borsig L, Varki NM, Varki A. P-selectin deficiency attenuates tumor growth and metastasis. *Proc Natl Acad Sci U S A*. 1998; 95:9325–30. [PubMed: 9689079]
22. Borsig L, Wong R, Feramisco J, Nadeau DR, Varki NM, Varki A. Heparin and cancer revisited: mechanistic connections involving platelets, P-selectin, carcinoma mucins, and tumor metastasis. *Proc Natl Acad Sci U S A*. 2001; 98:3352–7. [PubMed: 11248082]
23. Labelle M, Begum S, Hynes RO. Direct Signaling between Platelets and Cancer Cells Induces an Epithelial-Mesenchymal-Like Transition and Promotes Metastasis. *Cancer Cell*. 2011; 20:576–90. [PubMed: 22094253]
24. Gasic GJ, Gasic TB, Stewart CC. Antimetastatic effects associated with platelet reduction. *Proc Natl Acad Sci U S A*. 1968; 61:46–52. [PubMed: 5246932]
25. Pearlstein E, Ambrogio C, Karpatkin S. Effect of antiplatelet antibody on the development of pulmonary metastases following injection of CT26 colon adenocarcinoma, Lewis lung carcinoma, and B16 amelanotic melanoma tumor cells into mice. *Cancer Res*. 1984; 44:3884–7. [PubMed: 6744304]
26. Camerer E, Qazi AA, Duong DN, Cornelissen I, Advincula R, Coughlin SR. Platelets, protease-activated receptors, and fibrinogen in hematogenous metastasis. *Blood*. 2004; 104:397–401. [PubMed: 15031212]
27. Trousseau, A. *Clinique Medicale de l'Hotel-Dieu de Paris*. 2. Paris: JB Bailliere & Fils; 1865. Phlegmasia alba dolens; p. 654-712.

28. Lyman GH, Khorana AA. Cancer, clots and consensus: new understanding of an old problem. *J Clin Oncol.* 2009; 27:4821–6. [PubMed: 19752337]
29. Kopp HG, Placke T, Salih HR. Platelet-derived transforming growth factor-beta down-regulates NKG2D thereby inhibiting natural killer cell antitumor reactivity. *Cancer Res.* 2009; 69:7775–83. [PubMed: 19738039]
30. Gersuk GM, Westernmark B, Mohabeer AJ, Challita PM, Pattamakom S, Pattengale PK. Inhibition of human natural killer cell activity by platelet-derived growth factor (PDGF). III. Membrane binding studies and differential biological effect of recombinant PDGF isoforms. *Scand J Immunol.* 1991; 33:521–32. [PubMed: 1851574]
31. Placke T, Oergel M, Schaller M, Jung G, Rammensee HG, Kopp HG, et al. Platelet-derived MHC Class I confers a pseudo-normal phenotype to cancer cells that subverts the anti-tumor reactivity of natural killer immune cells. *Cancer Res.* 2011
32. Stott SL, Hsu CH, Tsukrov DI, Yu M, Miyamoto DT, Waltman BA, et al. Isolation of circulating tumor cells using a microvortex-generating herringbone-chip. *Proc Natl Acad Sci U S A.* 2010; 107:18392–7. [PubMed: 20930119]
33. Duda DG, Duyverman AM, Kohno M, Snuderl M, Steller EJ, Fukumura D, et al. Malignant cells facilitate lung metastasis by bringing their own soil. *Proc Natl Acad Sci U S A.* 2010; 107:21677–82. [PubMed: 21098274]
34. Al-Mehdi AB, Tozawa K, Fisher AB, Shientag L, Lee A, Muschel RJ. Intravascular origin of metastasis from the proliferation of endothelium-attached tumor cells: a new model for metastasis. *Nat Med.* 2000; 6:100–2. [PubMed: 10613833]
35. Orr FW, Wang HH. Tumor cell interactions with the microvasculature: a rate-limiting step in metastasis. *Surg Oncol Clin N Am.* 2001; 10:357–81. ix–x. [PubMed: 11382592]
36. Wang H, Fu W, Im JH, Zhou Z, Santoro SA, Iyer V, et al. Tumor cell alpha3beta1 integrin and vascular laminin-5 mediate pulmonary arrest and metastasis. *J Cell Biol.* 2004; 164:935–41. [PubMed: 15024036]
37. Brown DM, Ruoslahti E. Metadherin, a cell surface protein in breast tumors that mediates lung metastasis. *Cancer Cell.* 2004; 5:365–74. [PubMed: 15093543]
38. Hu G, Chong RA, Yang Q, Wei Y, Blanco MA, Li F, et al. MTDH activation by 8q22 genomic gain promotes chemoresistance and metastasis of poor-prognosis breast cancer. *Cancer Cell.* 2009; 15:9–20. [PubMed: 19111877]
39. Luzzi KJ, MacDonald IC, Schmidt EE, Kerkvliet N, Morris VL, Chambers AF, et al. Multistep nature of metastatic inefficiency: dormancy of solitary cells after successful extravasation and limited survival of early micrometastases. *Am J Pathol.* 1998; 153:865–73. [PubMed: 9736035]
40. Laubli H, Stevenson JL, Varki A, Varki NM, Borsig L. L-selectin facilitation of metastasis involves temporal induction of Fut7-dependent ligands at sites of tumor cell arrest. *Cancer Res.* 2006; 66:1536–42. [PubMed: 16452210]
41. Palumbo JS, Talmage KE, Massari JV, La Jeunesse CM, Flick MJ, Kombrinck KW, et al. Tumor cell-associated tissue factor and circulating hemostatic factors cooperate to increase metastatic potential through natural killer cell-dependent and-independent mechanisms. *Blood.* 2007; 110:133–41. [PubMed: 17371949]
42. Qian B, Deng Y, Im JH, Muschel RJ, Zou Y, Li J, et al. A distinct macrophage population mediates metastatic breast cancer cell extravasation, establishment and growth. *PLoS One.* 2009; 4:e6562. [PubMed: 19668347]
43. Kienast Y, von Baumgarten L, Fuhrmann M, Klinkert WE, Goldbrunner R, Herms J, et al. Real-time imaging reveals the single steps of brain metastasis formation. *Nat Med.* 2010; 16:116–22. [PubMed: 20023634]
44. Hiratsuka S, Goel S, Kamoun WS, Maru Y, Fukumura D, Duda DG, et al. Endothelial focal adhesion kinase mediates cancer cell homing to discrete regions of the lungs via E-selectin up-regulation. *Proc Natl Acad Sci U S A.* 2011; 108:3725–30. [PubMed: 21321210]
45. Kaplan RN, Riba RD, Zacharoulis S, Bramley AH, Vincent L, Costa C, et al. VEGFR1-positive haematopoietic bone marrow progenitors initiate the pre-metastatic niche. *Nature.* 2005; 438:820–7. [PubMed: 16341007]

46. Hiratsuka S, Watanabe A, Aburatani H, Maru Y. Tumour-mediated upregulation of chemoattractants and recruitment of myeloid cells predetermines lung metastasis. *Nat Cell Biol.* 2006; 8:1369–75. [PubMed: 17128264]
47. Hiratsuka S, Watanabe A, Sakurai Y, Akashi-Takamura S, Ishibashi S, Miyake K, et al. The S100A8-serum amyloid A3-TLR4 paracrine cascade establishes a pre-metastatic phase. *Nat Cell Biol.* 2008; 10:1349–55. [PubMed: 18820689]
48. Erler JT, Bennewith KL, Cox TR, Lang G, Bird D, Koong A, et al. Hypoxia-induced lysyl oxidase is a critical mediator of bone marrow cell recruitment to form the premetastatic niche. *Cancer Cell.* 2009; 15:35–44. [PubMed: 19111879]
49. Deng J, Liu Y, Lee H, Herrmann A, Zhang W, Zhang C, et al. S1PR1-STAT3 Signaling Is Crucial for Myeloid Cell Colonization at Future Metastatic Sites. *Cancer Cell.* 2012; 21:642–54. [PubMed: 22624714]
50. Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. *Nature.* 2008; 454:436–44. [PubMed: 18650914]
51. Qian BZ, Li J, Zhang H, Kitamura T, Zhang J, Campion LR, et al. CCL2 recruits inflammatory monocytes to facilitate breast-tumour metastasis. *Nature.* 2011; 475:222–5. [PubMed: 21654748]
52. Iwai K, Ishikura H, Kaji M, Sugiura H, Ishizu A, Takahashi C, et al. Importance of E-selectin (ELAM-1) and sialyl Lewis(a) in the adhesion of pancreatic carcinoma cells to activated endothelium. *Int J Cancer.* 1993; 54:972–7. [PubMed: 7687590]
53. Aigner S, Stoeber ZM, Fogel M, Weber E, Zarn J, Ruppert M, et al. CD24, a mucin-type glycoprotein, is a ligand for P-selectin on human tumor cells. *Blood.* 1997; 89:3385–95. [PubMed: 9129046]
54. Khatib AM, Auguste P, Fallavollita L, Wang N, Samani A, Kontogianna M, et al. Characterization of the host proinflammatory response to tumor cells during the initial stages of liver metastasis. *Am J Pathol.* 2005; 167:749–59. [PubMed: 16127154]
55. Laubli H, Spanaus KS, Borsig L. Selectin-mediated activation of endothelial cells induces expression of CCL5 and promotes metastasis through recruitment of monocytes. *Blood.* 2009; 114:4583–91. [PubMed: 19779041]
56. Massberg S, Konrad I, Schurzinger K, Lorenz M, Schneider S, Zohlnhoefer D, et al. Platelets secrete stromal cell-derived factor 1alpha and recruit bone marrow-derived progenitor cells to arterial thrombi in vivo. *J Exp Med.* 2006; 203:1221–33. [PubMed: 16618794]
57. Huo Y, Schober A, Forlow SB, Smith DF, Hyman MC, Jung S, et al. Circulating activated platelets exacerbate atherosclerosis in mice deficient in apolipoprotein E. *Nat Med.* 2003; 9:61–7. [PubMed: 12483207]
58. Ludwig RJ, Schultz JE, Boehncke WH, Podda M, Tandl C, Krombach F, et al. Activated, not resting, platelets increase leukocyte rolling in murine skin utilizing a distinct set of adhesion molecules. *J Invest Dermatol.* 2004; 122:830–6. [PubMed: 15086572]
59. Dole VS, Bergmeier W, Mitchell HA, Eichenberger SC, Wagner DD. Activated platelets induce Weibel-Palade-body secretion and leukocyte rolling in vivo: role of P-selectin. *Blood.* 2005; 106:2334–9. [PubMed: 15956287]
60. Diacovo TG, deFougerolles AR, Bainton DF, Springer TA. A functional integrin ligand on the surface of platelets: intercellular adhesion molecule-2. *J Clin Invest.* 1994; 94:1243–51. [PubMed: 8083366]
61. Weber C, Springer TA. Neutrophil accumulation on activated, surface-adherent platelets in flow is mediated by interaction of Mac-1 with fibrinogen bound to alphaIIb beta3 and stimulated by platelet-activating factor. *J Clin Invest.* 1997; 100:2085–93. [PubMed: 9329974]
62. Simon DI, Chen Z, Xu H, Li CQ, Dong J, McIntire LV, et al. Platelet glycoprotein Iba1 is a counterreceptor for the leukocyte integrin Mac-1 (CD11b/CD18). *J Exp Med.* 2000; 192:193–204. [PubMed: 10899906]
63. Santoso S, Sachs UJ, Kroll H, Linder M, Ruf A, Preissner KT, et al. The junctional adhesion molecule 3 (JAM-3) on human platelets is a counterreceptor for the leukocyte integrin Mac-1. *J Exp Med.* 2002; 196:679–91. [PubMed: 12208882]

64. Borsig L, Wong R, Hynes RO, Varki NM, Varki A. Synergistic effects of L- and P-selectin in facilitating tumor metastasis can involve non-mucin ligands and implicate leukocytes as enhancers of metastasis. *Proc Natl Acad Sci U S A*. 2002; 99:2193–8. [PubMed: 11854515]
65. Gil-Bernabe AM, Ferjancic S, Tlalka M, Zhao L, Allen PD, Im JH, et al. Recruitment of monocytes/macrophages by tissue factor-mediated coagulation is essential for metastatic cell survival and premetastatic niche establishment in mice. *Blood*. 2012; 119:3164–75. [PubMed: 22327225]
66. Huh SJ, Liang S, Sharma A, Dong C, Robertson GP. Transiently entrapped circulating tumor cells interact with neutrophils to facilitate lung metastasis development. *Cancer Res*. 2010; 70:6071–82. [PubMed: 20610626]
67. Acuff HB, Carter KJ, Fingleton B, Gorden DL, Matrisian LM. Matrix metalloproteinase-9 from bone marrow-derived cells contributes to survival but not growth of tumor cells in the lung microenvironment. *Cancer Res*. 2006; 66:259–66. [PubMed: 16397239]
68. Granot Z, Henke E, Comen EA, King TA, Norton L, Benezra R. Tumor entrained neutrophils inhibit seeding in the premetastatic lung. *Cancer Cell*. 2011; 20:300–14. [PubMed: 21907922]
69. Fridlender ZG, Sun J, Kim S, Kapoor V, Cheng G, Ling L, et al. Polarization of tumor-associated neutrophil phenotype by TGF-beta: "N1" versus "N2" TAN. *Cancer Cell*. 2009; 16:183–94. [PubMed: 19732719]
70. Gupta GP, Nguyen DX, Chiang AC, Bos PD, Kim JY, Nadal C, et al. Mediators of vascular remodelling co-opted for sequential steps in lung metastasis. *Nature*. 2007; 446:765–70. [PubMed: 17429393]
71. Bos PD, Zhang XH, Nadal C, Shu W, Gomis RR, Nguyen DX, et al. Genes that mediate breast cancer metastasis to the brain. *Nature*. 2009; 459:1005–9. [PubMed: 19421193]
72. Weis S, Cui J, Barnes L, Cheresh D. Endothelial barrier disruption by VEGF-mediated Src activity potentiates tumor cell extravasation and metastasis. *J Cell Biol*. 2004; 167:223–9. [PubMed: 15504909]
73. Padua D, Zhang XH, Wang Q, Nadal C, Gerald WL, Gomis RR, et al. TGFbeta primes breast tumors for lung metastasis seeding through angiopoietin-like 4. *Cell*. 2008; 133:66–77. [PubMed: 18394990]
74. Palframan RT, Jung S, Cheng G, Weninger W, Luo Y, Dorf M, et al. Inflammatory chemokine transport and presentation in HEV: a remote control mechanism for monocyte recruitment to lymph nodes in inflamed tissues. *J Exp Med*. 2001; 194:1361–73. [PubMed: 11696600]
75. Wolf MJ, Hoos A, Bauer J, Boettcher S, Knust M, Weber A, et al. Endothelial CCR2 signaling induced by colon carcinoma cells enables extravasation via the JAK2-Stat5 and p38MAPK pathway. *Cancer Cell*. 2012; 22:91–105. [PubMed: 22789541]
76. Chen Q, Zhang XH, Massague J. Macrophage Binding to Receptor VCAM-1 Transmits Survival Signals in Breast Cancer Cells that Invade the Lungs. *Cancer Cell*. 2011; 20:538–49. [PubMed: 22014578]
77. Oskarsson T, Acharyya S, Zhang XH, Vanharanta S, Tavazoie SF, Morris PG, et al. Breast cancer cells produce tenascin C as a metastatic niche component to colonize the lungs. *Nat Med*. 2011; 17:867–74. [PubMed: 21706029]
78. Malanchi I, Santamaria-Martinez A, Susanto E, Peng H, Lehr HA, Delaloye JF, et al. Interactions between cancer stem cells and their niche govern metastatic colonization. *Nature*. 2011; 481:85–9. [PubMed: 22158103]
79. Jinnin M, Ihn H, Asano Y, Yamane K, Trojanowska M, Tamaki K. Tenascin-C upregulation by transforming growth factor-beta in human dermal fibroblasts involves Smad3, Sp1, and Ets1. *Oncogene*. 2004; 23:1656–67. [PubMed: 15001984]
80. Oskarsson T, Massague J. Extracellular matrix players in metastatic niches. *EMBO J*. 2011
81. Steeg PS. Perspective: The right trials. *Nature*. 2012; 485:S58–9. [PubMed: 22648501]
82. Bernards R, Weinberg RA. A progression puzzle. *Nature*. 2002; 418:823. [PubMed: 12192390]
83. Husemann Y, Geigl JB, Schubert F, Musiani P, Meyer M, Burghart E, et al. Systemic spread is an early step in breast cancer. *Cancer Cell*. 2008; 13:58–68. [PubMed: 18167340]
84. Hoover HC Jr, Ketcham AS. Metastasis of metastases. *Am J Surg*. 1975; 130:405–11. [PubMed: 1166931]

85. Fidler IJ, Nicolson GL. Fate of recirculating B16 melanoma metastatic variant cells in parabiotic syngeneic recipients. *J Natl Cancer Inst.* 1977; 58:1867–72. [PubMed: 864765]
86. Kim MY, Oskarsson T, Acharyya S, Nguyen DX, Zhang XH, Norton L, et al. Tumor self-seeding by circulating cancer cells. *Cell.* 2009; 139:1315–26. [PubMed: 20064377]
87. Brown DC, Purushotham AD, Birnie GD, George WD. Detection of intraoperative tumor cell dissemination in patients with breast cancer by use of reverse transcription and polymerase chain reaction. *Surgery.* 1995; 117:95–101. [PubMed: 7809843]
88. Weitz J, Kienle P, Lacroix J, Willeke F, Benner A, Lehnert T, et al. Dissemination of tumor cells in patients undergoing surgery for colorectal cancer. *Clin Cancer Res.* 1998; 4:343–8. [PubMed: 9516921]
89. Lebeau B, Chastang C, Brechot JM, Capron F, Dautzenberg B, Delaisements C, et al. Subcutaneous heparin treatment increases survival in small cell lung cancer “Petites Cellules” Group. *Cancer.* 1994; 74:38–45. [PubMed: 8004580]
90. Kakkar AK, Levine MN, Kadziola Z, Lemoine NR, Low V, Patel HK, et al. Low molecular weight heparin, therapy with dalteparin, and survival in advanced cancer: the fragmin advanced malignancy outcome study (FAMOUS). *J Clin Oncol.* 2004; 22:1944–8. [PubMed: 15143088]
91. Klerk CP, Smorenburg SM, Otten HM, Lensing AW, Prins MH, Piovella F, et al. The effect of low molecular weight heparin on survival in patients with advanced malignancy. *J Clin Oncol.* 2005; 23:2130–5. [PubMed: 15699479]
92. Stevenson JL, Choi SH, Varki A. Differential metastasis inhibition by clinically relevant levels of heparins--correlation with selectin inhibition, not antithrombotic activity. *Clin Cancer Res.* 2005; 11:7003–11. [PubMed: 16203794]
93. Bendas G, Borsig L. Cancer cell adhesion and metastasis: selectins, integrins, and the inhibitory potential of heparins. *Int J Cell Biol.* 2012; 2012:676731. [PubMed: 22505933]
94. Nierodzic ML, Klepfish A, Karpatkin S. Role of platelets, thrombin, integrin IIb-IIIa, fibronectin and von Willebrand factor on tumor adhesion in vitro and metastasis in vivo. *Thromb Haemost.* 1995; 74:282–90. [PubMed: 8578473]
95. Okahara H, Yagita H, Miyake K, Okumura K. Involvement of very late activation antigen 4 (VLA-4) and vascular cell adhesion molecule 1 (VCAM-1) in tumor necrosis factor alpha enhancement of experimental metastasis. *Cancer Res.* 1994; 54:3233–6. [PubMed: 7515767]
96. Goodman SL, Picard M. Integrins as therapeutic targets. *Trends Pharmacol Sci.* 2012; 33:405–12. [PubMed: 22633092]
97. Chen Q, Massague J. Molecular pathways: VCAM-1 as a potential therapeutic target in metastasis. *Clin Cancer Res.* 2012

Significance

Metastasis is a function not only of tumor cells but also involves cooperative interactions of those cells with normal cells of the body, in particular platelets and leukocytes. These other cell types alter the behavior of the tumor cells themselves and of endothelial cells lining the vasculature and assist in tumor cell arrest and extravasation at sites of metastasis and subsequently in the establishment of tumor cells in the early metastatic niche. A better understanding of the important role that these contact and paracrine interactions play during metastasis will offer new opportunities for therapeutic intervention.

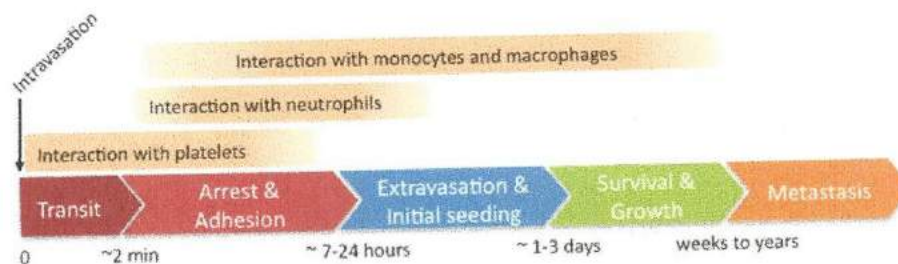


Figure 1. Temporal dynamics of host-tumor cell interactions during the early steps of the metastatic cascade

Tumor cells intravasate, rapidly transit through the circulation, and arrest in the vasculature of a secondary organ, generally within a few minutes. During this period, platelets form aggregates around CTCs or arrested tumor cells. Neutrophils also interact with tumor cells within the first day. Seven to 48 hours after tail-vein injection of tumor cells, monocytes/macrophages are also recruited to their vicinity. Extravasation typically takes place within the first 1–3 days after initial arrest. By that time, most tumor cells have exited the bloodstream and seeded into the stroma of the secondary site and additional myeloid cells are recruited to this initial metastatic niche. The tumor cells may reinitiate growth to form metastases within a few weeks. Alternatively, tumor cells can survive and stay dormant for a long period before reinitiating growth and thus form clinically relevant metastases only months or years later. Overall, only few cells successfully complete the metastatic cascade and give rise to overt metastases. So far, most studies of host-tumor cell interactions during metastasis have been performed at single time points arbitrarily chosen by investigators while only few studies have examined the temporal recruitment of host cells by real-time imaging or sampling at multiple time points. This scheme is thus a tentative summary of many independent studies reporting experimental observations at different time points and obtained with different model systems.

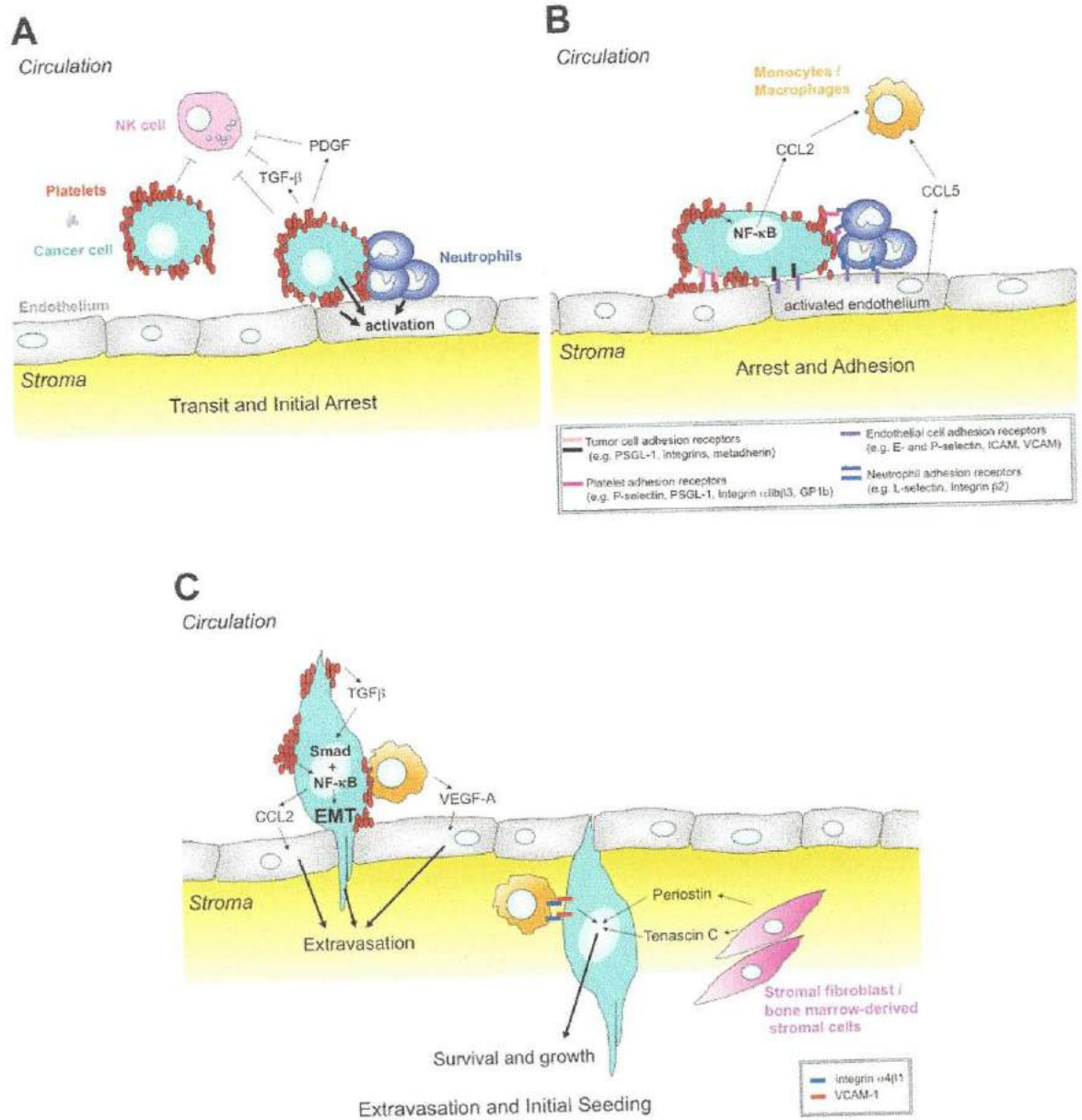


Figure 2. Examples of host-tumor cell interactions during the arrest and extravasation steps of the metastatic cascade

A. Upon entry into the blood circulation, tumor cells become exposed to interactions with various blood cells. Platelet-tumor cell interactions may occur soon after the entry of tumor cells into the circulation, when they are still circulating, or very early upon initial arrest. Overall, the interaction of platelets with tumor cells has various prometastatic functions during the vascular phase of the metastatic cascade. Early on, the formation of a platelet-rich thrombus around tumor cells protects tumor cells from shear stress and against lysis by NK cells. The formation of platelet microthrombi around tumor cells may also favor arrest and adhesion to the endothelium. Simultaneously, neutrophils may be recruited to the platelet-tumor cell aggregates, and participate in endothelial cell activation.

B. Cytokines and chemokines secreted by platelets, the tumor cells, neutrophils and the activated endothelium promote the recruitment of monocytes, which, in concert with the

other cells already present in this “niche”, further activate the endothelium and enhance tumor cell extravasation.

C. Direct signaling between platelets and tumor cells also contributes to extravasation by inducing a more invasive pro-metastatic phenotype in tumor cells and probably also through effects on the endothelium. The recruited BMDCs and other host cells such as fibroblasts then contribute to the remodeling of the microenvironment to further support initial tumor cell survival and growth in the tissue parenchyma, eventually leading to the formation of overt metastases.